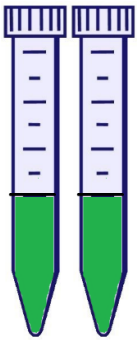
Transformation of 1.6 to synechocystis

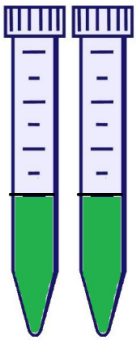
- Use cultures that say: use for transformation
- Measure and note down OD at **730nm.** Blank with MQ water is fine. Use **1 mL** in cuvette, if above linear range: use that sample to dilute with MQ water in another cuvette. Don't take more than 1 mL from the culture for OD_{730nm} measurements

Protocol:

- Centrifuge 2x 10 mL of healthy cells (both wild type and JA06) at an OD_{730nm} of ~ 0.7 at 4000x*g* for 6 min



WT 10 ml x2



JA06 10 ml x2

- Wash pellet twice in BG-11
- Resuspend the cell pellet in **200 µL** of BG-11 in a 2 ml eppendorf tube
- Add **1 µg** of target plasmid to 1 of the WT and 1 of the JA06. The other tube will be used as negative control (without added plasmid).
- Incubate the tube in light for 5 hours at 30 °C. Fasten the tube lying down on the shaker in the 30-degree room so it receives light and shaking.
- Transfer the 200 µL to a shake flask containing 4 ml BG-11 and grow 18-24 h
- Plate cells on BG-11 plates with appropriate antibiotics
    - **1.6: BG-11 + kanamycin + L-glutamine (5 mM)**